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# Impact of Salinity Stress on Seed Quality Parameters and its Mitigation through Seed Quality Enhancement in Blackgram Seeds (*Vigna mungo* L.)

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ABSTRACT: The Purpose of this study was to determine the impact of salinity stress on seed quality parameters and its mitigation through seed quality enhancement in blackgram seeds (*Vigna mungo* L.). The lab experiment was conducted during 2020-2021 in seed testing laboratory Department of Genetics and Plant Breeding, NAI, SHUATS, Prayagraj, Uttar Pradesh. Salinity stress on blackgram seeds on enhancement of seed quality parameters. The were given a treatment Panchagaya (3% and 6%), Bejamrita (25% and 50%), Coconut water (8% and 10%), KNO<sub>3</sub> (3grm and 6gm) in seed quality parameters under the various Salinity Stress condition of with three levels S<sub>0</sub>, S<sub>1</sub> and S<sub>2</sub> (0 mM, 100 mM, 150mM) with Shaker –II variety of Blackgram. The result revealed that treatments recorded significant variation with the untreated control. The treatment combination T<sub>8</sub>S<sub>0</sub> of 6gm KNO<sub>3</sub> for 12 hours at 0 mM NaCl performed well with seedling. Germination of Percentage 95.50%, Shoot length as 10.025 cm as follows as Root length 12.788 cm, Seedling length 22.803 cm, Shoot-Root ratio 0.684, Fresh weight 2.600 g, Dry weight 0.425 g, Seedling Vigour index–II 40.675, showed these supremacy on increased activity which protect the plant under salinity condition.

Keywords: Black gram, Organic priming, Inorganic priming, Salinity Stress.

## **INTRODUCTION**

Black gram (Vigna mungo L.) belonging to leguminoseae family, with chromosomal number 2n=2x=22 and is an essential nutritious pulse crop with a special place in Indian agriculture. Self-pollinated (Autogamy), Vigna mungo probably originated in South Asia. In India, Blackgram is commercially produced in Maharashtra, Andhra Pradesh, U.P., Madhya Pradesh, Tamilnadu, Rajasthan, Gujarat, Karnataka, Jharkhand and West Bengal. Urd bean residues contain about 10-11% moisture, 24% protein, 1.4-1.6% fat, 3.2% minerals, 16% dietary fibers, 59.69-63.0% carbohydrates, 154mg/100g calcium, 385mg/100g phosphorus, 9.1mg/100g iron, 347k.cal/100g calorific value, it has the highest concentration of phosphoric acid among pulses (5-6%). (Source: DPD, Govt, of India, pulses in India: Ritrospect and prospects, 2018). Seed priming is a pre-sowing method for affecting seed development by alternating pre-germination metabolic activity prior to radicle emergence, which improves fast, uniform emergence and plant performance high vigour and better yields (Mc Donald, 2000).

It's also a natural disinfectant and pest repellent, and it's one of the main elements of Panchagavya (an organic crop booster prepared and sprayed by Indian farmers) (Tharmaraj *et al.*, 2011). It is used as a foliar spray, soil application along with irrigation, as well as seed treatment (Natarajan, 2002). Bejamrita is a naturally available preservative based on seed treatment that protects the crop against damaging soil and seed borne diseases during the early stages of germination and establishment .In India, using bijamritha is a common practice. Organic manures are formed to contain necessary nutrient in previous experiments, resulting in enhanced crop growth and production (Vyankatrao, 2017). In that mostly coconut water, are used for seed priming. These plants contain compounds that cause pre-germinative phenomena, reducing germination time and enhancing seedling vigour. Low amounts of KNO<sub>3</sub> have been shown in previous research to help mitigate the negative effects of NaCl on seed germination in some grass species (Neid and Biesboer 2005). Because K+ is involved in a variety of plant activations. The K+/Na+ ratio has been suggested as useful biomarker for Wheat salinity tolerance, (Zheng et al., 2008).

Salinity can affect germination by producing an osmotic potential that inhibits water absorption or by the harmful effects of ions on embryo survival (Houle *et al.*, 2001). Salts absorb and hold water to the point that is no longer readily accessible in the soil, resulting in a rise in the osmotic pressure of the soil solution. Salt stress can result in considerable decreases in germination rate and final germination percentage,

which can lead to uneven stand establishment and lower yield.

Salt stress hampered the agricultural productivity by lowering yield of various crops in arid and semi-arid region of the world. Salt stress greatly influenced the germination of blackgram seed. The germination rate determines the crop productivity by optimizing the germination factor.

#### MATERIAL AND METHODS

The current research was conducted during the year 2020-2021 at Department of Genetics and Plant Breeding, NAI, SHUATS (Uttar Pradesh). The lab experiment was evaluated using CRD (Completely Randomized Design) three salinity levels goes through

laboratory conditions. Seeds treated with  $T_0$ -Control,  $T_1$ -Panchagavya (3%),  $T_2$ - Panchagavya (6%),  $T_3$ -Bijamrita (25%),  $T_4$ - Bijamrita (50%),  $T_5$ - Coconut water (8%),  $T_6$ - Coconut water (10%),  $T_7$ - KNO<sub>3</sub> (3g) and  $T_8$ - KNO<sub>3</sub> (6g) 12 hours of soaking after that, primed seeds were allowed to dry to their original moisture content in the shade to evalveture the parameters at three different salt levels: 0 mM, 100mM and 150 mM of NaCl. Germination percentage, Shoot length, Root length, Seedling length, Shoot-root ratio, Seedling fresh weight, Seedling dry weight, Seedling Vigour Index- I and seedling vigour index – II are some of the seed quality characteristics.

Table 1: Analysis of variance for different seedling growth parameters in blackgram under salinity stress.

Sr. No.		Mean sum of square						
	Characters	Salinity stress (df=2)	Treatments (df=8)	Interactions (df=16)	Error (df=81)			
1.	Germination %	6,388.731*	129.475*	24.836*	1.284			
2.	Shoot length	231.122*	5.384*	$2.622^{*}$	0.979			
3.	Root length	863.941*	3.741*	$1.680^{*}$	0.883			
4.	Seedling length	1,957.355*	17.103*	4.193*	1.763			
5.	Shoot-Root ratio	10.609*	0.808*	0.923*	0.352			
б.	Seedling fresh weight	18.405*	0.764*	0.046*	0.021			
7.	Seedling dry weight	0.532*	0.059*	0.010*	0.005			
8.	Vigour index – I	20,631,521.604*	256,154.669*	30,000.586*	16,845.577			
9.	Vigour index – II	5,575.030*	553.407 <sup>*</sup>	90.657*	43.114			

\* Significant at 5% level of significance.

#### Methodology:

**Germination%:** At the end of 7<sup>th</sup> day of sowing, the no. of normal seedling in each replication was conducted and the germination was calculated and expressed in percentage.

Germination (%) =  $\frac{\text{No. of seeds germinated}}{\text{No. of seeds put for germination}} \times 100$ 

**Shoot (Plumule) length (cm):** On the seventh day after the germination test, ten normal seedlings will be chosen at random from each replication in each salinity level and treatment. From the tip of the main leafs to the base of the hypocotyl, the length of the shoot will be measured.

**Root length (cm):** The root length will be measured using the same 10 seedlings that were used to measure the shoot length. The length of the root from the tip of the primary root to the base of the hypocotyl.

**Seeding length (cm):** The sum of shoot and root length will be considered as seedling length.

**Shoot-Root ratio:** On the seventh day after germination, ten normal seedlings will be randomly picked from each replication in each salinity level, and each treatments ratio of shoot length to root length will be assessed separately.

**Seedling Fresh Weight (g):** For recording seedling fresh weight are weighed with help of electronic balance.

**Seedling Dry Weight (g):** Seedlings are dried for 24 hours in an 800°F hot air oven to record their dry weight.

Seedling Vigour Index-I: germination Percentage  $\times$  Total seedling length as cm.

**Seedling Vigour Index- II:** germination Percentage  $\times$  seedling dry weight as g.



Fig. 1. Placing the seeds for Germination using between paper method.



Fig. 2. Keeping for germination in germination room.

### **RESULT AND DISCUSSION**

According to the findings, all of the studied traits where affected by the treatments and salinity stress and all seed quality parameters, such as germination percentage, shoot length, root length, seedling length, shoot-root ratio, seedling fresh weight, seedling dry weight, seedling vigour index-I and seedling vigour index-II showed a completely significant difference between control and primed seeds. In Table 2, the data penetrating that significantly higher germination % in (S<sub>0</sub>-95.50), (S<sub>1</sub>-94.00), (S<sub>2</sub>-77.00) reported in T<sub>8</sub> 6g of KNO<sub>3</sub> followed by T<sub>7</sub> 3g of KNO<sub>3</sub> (S<sub>0</sub>-95.00, (S<sub>1</sub>-93.25), (S<sub>2</sub>-75.25) Kaya *et al.*, (2006) reported by positive effects of hydro priming & priming along with KNO<sub>3</sub> Bocian and Holubowicz (2008) were also reported that priming with KNO<sub>3</sub> enhanced seed germination of tomato.

Table 2: Comparison between overall mean of different treatments under different salinity levels in urd bean.

Sr. No.	Treatments	Salinity levels	Germination %	Shoot length (cm)	Root length (cm)	Seedling length (cm)	Shoot- Root Ratio	Seedling Fresh Weight (g)	Seedling Dry Weight (g)	Vigour Index – I	Vigour Index - II
1.	To	S <sub>0</sub>	89.500	8.452	12.210	20.663	1.039	1.875	0.200	1,849.450	17.925
		<b>S</b> 1	88.250	6.143	5.353	11.495	1.828	0.990	0.098	1,014.113	8.608
		<b>S</b> <sub>2</sub>	60.250	4.070	1.200	5.270	3.137	0.555	0.055	317.845	3.318
2.	T <sub>1</sub>	S <sub>0</sub>	91.250	9.430	12.753	22.183	0.746	1.950	0.250	2,024.550	22.875
		<b>S</b> <sub>1</sub>	89.500	6.273	4.375	10.648	1.464	1.020	0.100	953.388	8.950
		<b>S</b> <sub>2</sub>	61.250	4.550	1.688	6.238	2.658	0.630	0.068	382.920	4.138
3.	T <sub>2</sub>	S <sub>0</sub>	92.250	8.215	12.093	20.308	0.952	2.000	0.300	1,872.725	27.725
		<b>S</b> <sub>1</sub>	90.500	5.685	4.588	10.273	1.311	1.200	0.150	930.983	13.600
		<b>S</b> <sub>2</sub>	64.000	4.358	2.275	6.633	1.914	0.723	0.073	424.923	4.643
4.		S <sub>0</sub>	93.000	9.465	11.255	20.720	1.089	2.175	0.325	1,925.960	30.300
	-	<b>S</b> <sub>1</sub>	90.750	7.168	5.700	12.868	1.263	1.200	0.175	1,167.900	15.925
	$T_3$	<b>S</b> <sub>2</sub>	69.250	3.735	2.068	5.803	1.843	0.820	0.075	403.355	5.198
		S <sub>0</sub>	94.000	9.921	11.420	21.341	0.884	2.350	0.325	2,005.877	30.675
-	T <sub>4</sub>	<b>S</b> 1	91.250	7.215	4.623	11.838	1.705	1.325	0.200	1,081.015	18.300
5.		<b>S</b> <sub>2</sub>	70.750	3.563	2.605	6.168	1.420	0.893	0.085	436.193	6.015
	T <sub>5</sub>	S <sub>0</sub>	94.250	9.445	11.970	21.415	0.788	2.400	0.350	2,018.022	33.000
		<b>S</b> 1	92.000	7.453	5.675	13.128	1.324	1.550	0.275	1,207.935	25.325
6.		<b>S</b> <sub>2</sub>	72.250	4.173	1.875	6.048	2.731	0.915	0.090	437.350	6.510
7.	T <sub>6</sub>	S <sub>0</sub>	94.750	9.435	10.143	19.578	0.937	2.425	0.375	1,856.450	35.575
		<b>S</b> 1	92.750	7.505	5.058	12.563	1.158	1.675	0.350	1,165.845	32.500
		<b>S</b> <sub>2</sub>	73.750	1.820	1.950	3.770	1.293	0.943	0.093	277.690	6.830
8.	<b>T</b> <sub>7</sub>	S <sub>0</sub>	95.000	9.475	12.168	21.643	0.780	2.500	0.400	2,056.035	38.050
		<b>S</b> <sub>1</sub>	93.250	7.743	5.508	13.250	0.938	1.800	0.375	1,235.500	35.000
		<b>S</b> <sub>2</sub>	75.250	5.617	3.250	8.868	1.526	0.975	0.095	743.135	7.153
9.	T <sub>8</sub>	S <sub>0</sub>	95.500	10.025	12.788	22.803	0.684	2.600	0.425	2,281.982	40.675
		S <sub>1</sub>	94.000	7.925	6.558	14.483	0.716	1.950	0.400	1,361.153	37.675
		S <sub>2</sub>	77.000	6.430	3.725	10.155	0.731	1.023	0.150	888.475	11.650
	Grand M	lean	84.648	6.863	6.476	13.339	1.378	1.499	0.217	1,197.07	19.561
	SE (d)	(S)	0.267	0.233	0.221	0.313	0.140	0.034	0.016	30.592	1.548
		(T)	0.463	0.404	0.384	0.542	0.242	0.059	0.028	52.987	2.681
	SE (M)	(S)	0.189	0.165	0.157	0.221	0.099	0.024	0.011	21.632	1.094
		(T)	0.327	0.286	0.271	0.383	0.171	0.042	0.020	37.467	1.895
	CD @5%	(S)	0.532	0.465	0.442	0.624	0.279	0.068	0.032	60.982	3.085
		(T)	0.922	0.805	0.765	1.080	0.483	0.117	0.056	105.624	5.344

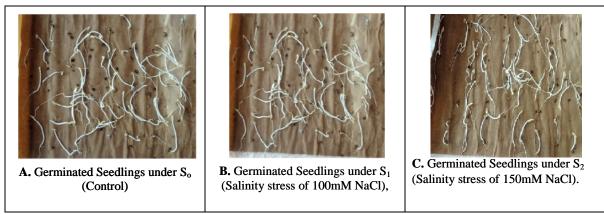
The germination % of primed with KNO<sub>3</sub> were more than unprimed seeds by Afkari (2010). Significantly higher shoot length were recorded in T<sub>8</sub> 6g of KNO<sub>3</sub> (S<sub>0</sub>-10.02), (S<sub>1</sub>-7.92) and (S<sub>2</sub>-6.43) followed by T<sub>7</sub> 3g of KNO<sub>3</sub> (S<sub>0</sub>-9.47), (S<sub>1</sub>-7.74) and (S<sub>2</sub>-5.61). Farooq *et al.*, (2015) reported that reduction in shoot length might be caused by a reduced number of elongated cells and the reduced right of cell elongation and this happen due to lower transport rate of essential ions like NO<sub>3</sub>- due to salinity that reduce the N compounds and increased Na+ in plant under high salinity (Hamid *et al.*, 2008). Significantly higher radicle length in T<sub>8</sub> 6g of KNO<sub>3</sub> (S<sub>0</sub>-12.78), (S<sub>1</sub>-6.58) and (S<sub>2</sub>-3.72) followed by T<sub>7</sub> 3g of KNO<sub>3</sub> (S<sub>0</sub>-12.16), (S<sub>1</sub>-5.50) and (S<sub>2</sub>-3.25) Kaya *et al.*, 2006 found that the difference in radicle length between priming treatments at the salinity levels of 4 and 8 dsm i.e., primed seeds showed a significant superiority over unprimed once.



Fig. 3. (A). Recording the Seedling length using Purity work boa. (B). Recording fresh weight of the seedling on the day final count.

Significantly highest shoot-ratio in  $T_0$  control (S<sub>0</sub>-1.03),  $(S_1-1.82)$  and  $(S_2-3.13)$  followed by  $(S_0-0.68)$ ,  $(S_1-0.71)$ and  $(S_2-0.73)$  due to the application of saline water by Reddy et al., (2013), the shoot and root length of blackgram progressively decreased. Significantly higher fresh weight were recorded T<sub>8</sub> 6g of KNO<sub>3</sub> (S<sub>0</sub>-2.60),  $(S_1-1.95)$  and  $(S_2-1.02)$  followed by  $T_7$  3g of KNO<sub>3</sub> (S<sub>0</sub>-2.50), (S<sub>1</sub>-1.80) and (S<sub>2</sub>-0.97) Aloui et al., (2014), hypothesised that the decreases in seedling fresh weight was related to a decrease in seedling water absorption for a lowering osmotic potential under salinity condition. Significantly higher dry weight of seedling were recorded in T<sub>8</sub>- 6g of KNO<sub>3</sub> (S<sub>0</sub>- 0.42), (S<sub>1</sub>-0.40), (S<sub>2</sub>-0.15) followed by  $T_7$  3g of KNO<sub>3</sub> (S<sub>0</sub>-0.40), (S<sub>1</sub>-0.37), (S<sub>2</sub>-0.09) Islam et al., (2011) were revealed that the in comparison to the control, stem dry

weight dropped as salt increased. Seedling vigour index-I was much greater reported in T<sub>8</sub> 6g of KNO<sub>3</sub> (S<sub>0</sub>-2, 281.98), (S<sub>1</sub>-1361.15), (S<sub>2</sub>-888.47) followed by T<sub>7</sub> 3g KNO<sub>3</sub> (S<sub>0</sub>-2056.03), (S<sub>1</sub>-1235.50), (S<sub>2</sub>-743.13). According to Cokkizgin, 2012, the seedling vigour index raise as the NaCl content decreased, indicating that higher NaCl concentrations damage seedling. Khajeh-Hosseini et al., (2003), in soybean, Saroj and Soumana (2014), in Vigna Spp. Found similar trends. Significantly higher seedling vigour index - II were recorded in T<sub>8</sub> 6g of KNO<sub>3</sub> (S<sub>0</sub>-40.67), (S<sub>1</sub>-37.67), (S<sub>2</sub>-11.65) followed by T<sub>7</sub> 3g KNO<sub>3</sub> (S<sub>0</sub>-38.05), (S<sub>1</sub>-35.00), (S<sub>2</sub>-7.15) Hassan et al., (2016) in wheat the cellular toxicity of NaCl in seedlings and reduction of chlorophyll content under saline condition which may also effects the seedling vigour index-II.



#### CONCLUSION

From the present investigation it is concluded that the seed treatment enhanced all the seed quality parameters under salinity stress in blackgram seeds when compared to control. It was also revealed that seed treatment with 6g of  $KNO_3$  is easy and effective approach to enhance the seed quality parameters under salinity stress.

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### FUTURE SCOPE

The further investigation needs to conduct for the field trial in the research farm using salt effected soils in the different region of the country as the balckgram variety used in this study Shekhar-II which is widely recommended in the different states in the India.

### REFERENCES

- Afkari, A. (2010). The effects of NaCl priming on salt tolerance in sunflower germination and seedling grown under salinity conditions. *Afri. J. Biotech.*, 9(12): 1764-1770.
- Aloui, H., Souguir, M., & Hannachi, C. (2014). Effect of salt stress (NaCl) on germination and early seedling parameters of three pepper cultivars (*Capsicum* annuum L.). J. Stress physiol. Biochem., 10(1): 14-25.
- Bocian, S.and R. Holubowicz, (2008). Effect of different ways of priming tomato (*Lycopersicon esculentum* Mill.) seeds on their quality. *Pol*. J. Nature Science, 23(4): 729-739.
- Farooq, M., Hussain, M., Wakeel, A., & Siddique K. H. M., (2015). Salt stress in maize effects, resistance mechanisms, and management, a review. Agron Sustain Dev, 35: 461-481.
- Hamid, M., Ashraf, M. Y., & Arashad, M. (2008). Influence of salicylic acid seed priming on growth and some biochemical attributes in wheat grown under saline conditions. *Pak. J. Bot.*, 40: 361-367.
- Houle, G. L., Morel, L., Reynolds, C. E., & Seigel, J. (2001). The effect of salinity on different developmental stages of an endemic annual plant, *Aster laurentianus*. *Am. J. Bot.*, 88: 62-67.
- Hasan, M. A., Alom, R., Islam, M. R., & Wang, Q. F. (2016). Germination characters and early seedling growth of

wheat (*Triticum aestivum* L.) genotypes under salt stress conditions. *Crop Sci.*, *Biotech.*, *19*(5): 383-392.

- Islam, M. S., Akhter, M.M., El Sabagh, A., Liu, L. Y., Nguyen, N. T., Ueda, A., Masaoka, Y., & Saneoka, H. (2011). Comparative studies on growth and physiological response to saline and alkaline stress of foxtail millet (*Setaria italica* L.) and Proso millet (*Panicum miliaceum* L.) Aust. J. Crop science, 5(10): 1269-1277.
- Kaya, M. D., Okçu, G., Atak, M., Cıkılı, Y., & Kolsarıcı, Ö. (2006). Seed treatments to overcome salt and drought stress during germination in sunflower (*Helianthus* annuus L.). European Journal of Agronomy, 24(4): 291-295.
- Khajeh-Hosseini, M., Powell, A. A., & Bingham, I. J. (2003). The interaction between salinity stress and seed vigour during germination of soybean seeds. *Seed science* and technology, 31(3): 715-725.
- Mc Donald, M. B. (2000). Seed priming. In: Seed technology and its biological basis. Black, M., Bewley J.D. (Eds). Sheffield Academic press, Sheffield, UK pp 287-325.157 Toklu F/ Not Bot Horti. Agrobo., 43(1): 153-158.
- Natarajan, K. (2002). Panchagavya A Manual. Other Indian press, Mapusa, Goa, India, pp: 333.
- Neid, S. L., & Biesboer, D. D. (2005). Alleviation of salt induced stress on seed emergence using soil additives in a green house. *Plant Soil*, 268, 303-307.
- Reddy, A. K., Yugandhar, P., & Savitharamma, N. (2013). Calcium chloride ameliorated sodium chloride salt stress in seedling growth of black gram (*Vigna mungo* L. Hepper). *International. Journal Advanced Science* and Technology, Research, 5(3): 151-161.
- Saroj, M., & Soumana, D. (2014). Salt stress induced changes in growth of germinating seeds of *Vigna mungo* (L.) Hepper and *Vigna aconitifolia* (Jacq.) Marechal. *IOSRJ. Agric.Vet. Sci.*, 7 (4): 44-48.
- Zheng, Y., Jia, A., Ning, T., Xu, J., Li, Z. & Jian, G. (2008). Potassium nitrate application alleviates sodium chloride stress in water wheat cultivars differing in salt tolerance. J. P 1. Physio1., 165(14): 1455-1465.
- Tharmaraj, K, Ganesh, P, Suresh kumar, R, Anandan, A. & Kolanjinathan, K. (2011). A Critical Review on Panchagavya – A Boon Plant Growth. Int. J. Pharm. Biol. Arch., 2(6): 1611-1614.
- Vyankatrao, N. P. (2017). Conversion of *Parthenium* hysterophorus L. Weed to compost and vermicompost. *Biosci Discov.*, 8(3): 619-627.

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